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Publisher *Taylor & Francis*

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Journal of Liquid Chromatography & Related Technologies

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713597273>

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To cite this Article Zennie, T. M.(1984) 'Instrumental HPTLC of Aflatoxins: Fluorescence Enhancement by Corn Free Fatty Acids', *Journal of Liquid Chromatography & Related Technologies*, 7: 7, 1383 – 1391

To link to this Article: DOI: 10.1080/01483918408074052

URL: <http://dx.doi.org/10.1080/01483918408074052>

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**INSTRUMENTAL HPTLC OF AFLATOXINS: FLUORESCENCE ENHANCEMENT
BY CORN FREE FATTY ACIDS**

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ABSTRACT

Greater than 100% recoveries using instrumental HPTLC were observed for aflatoxin (AFT) analysis in spiked corn samples. I₂ detection of spots over-laying AFT B₁ and B₂ were identified by GLC as C₁₆-C₁₈ free fatty acids (FFA). These FFA were found to enhance the fluorescence of AFT B₁ from 13.7% to 35.7% greater than controls resulting in >100% recoveries. The inclusion of glacial acetic acid in the TLC mobile phase resulted in an increased mobility of the FFA which eliminated the positive interference on AFT fluorescence. Recoveries using the modified developing solvent then gave values in acceptable ranges.

INTRODUCTION

During the development of a method to analyze for aflatoxins in small samples of corn and corn dusts (1.0 g-0.01 g), a positive error in B₁ and B₂ values was consistently observed during aflatoxin (AFT) spiking and recovery studies using instrumental HPTLC with fluorescence detection.

An actual example would consist of three 1.0 g samples of ground corn spiked with 50 ppb AFT B₁ and 10 ppb AFT B₂. After appropriate extraction and clean-up according to Zennie *et al.* (1), the AFT 's were quantified on HPTLC plates. Recovery would range from 56 to 83 ppb for B₁ and 12 and 14 ppb for B₂. These results show a 112 to 166% recovery for B₁ and a 120 to 140% recovery for B₂.

The most obvious cause for the increased fluorescence enhancement would be corn naturally contaminated with AFT; this however was ruled out by analyzing the unspiked corn and finding no discernable amounts of AFT 's. Inspection of the thin-layer plate under long-wave UV light revealed plainly visible AFT spots from both AFT standards and AFT spiked samples with no obvious overlapping or interfering fluorescent background material at or near the AFT 's. However, when the AFT areas were marked with pencil under UV light and then the plate placed in an I₂ tank, large areas which directly overlaid the AFT spots became visible. These were large tear shaped spots which had R_f values slightly ahead of AFT B₁ and B₂, but which tailed over the AFT B₁ and B₂ areas. Removal of this interference for accurate AFT determination is warranted.

EXPERIMENTAL

Column chromatography and preparatory TLC of corn extracts free of AFT resulted in the isolation of the interfering spot. The IR spectrum of the isolated material (neat on NaCl plates) gave absorptions at 1720 cm⁻¹ and a large broad band between 3400 cm⁻¹

and 2500 cm^{-1} which are suggestive of a carboxylic acid. Additional strong absorptions at 2920 cm^{-1} and 2850 cm^{-1} and another at 720 cm^{-1} denoted a long chain fatty acid. A stretching vibration at 3010 cm^{-1} revealed unsaturation. The material was esterified using BF_3 in methanol and run on GLC and positively identified as a typical mixture of long chain fatty acids seen in corn, i.e., palmitic, stearic, oleic, linoleic, and linolenic acid.

All HPTLC experiments were performed using $10\text{ cm} \times 10\text{ cm}$. HPTLC plates (E. Merck) developed in an unlined tank. The developing solvents for all experiments were either solvent A (8% acetone in chloroform) or solvent B (chloroform, acetone, glacial acetic acid [92+8+1]). The fluorescence enhancement studies and the free fatty acid (FFA) R_f determinations, were done by over-spotting the AFT on various concentrations of FFA. All AFT measurements were done on a Camag variable wave-length densitometer in fluorescence mode with a Hewlett Packard 3390A reporting integrator. Light source was a mercury lamp with excitation at 365 nm using a 400 nm cut off filter. Scanning slit width was $5\text{ mm} \times 0.3\text{ mm}$ with a 0.5 mm/sec scan speed.

Samples of ground corn (1.0 g) were extracted with 25 ml CHCl_3 , 1.0 g celite and 1 ml H_2O in a 125 ml Erlenmeyer flask with 30 min of shaking. This method of extraction results in approximately 10 mg of extractables for 1.0 g corn.

For the AFT recovery experiment an extract from 1.0 g of AFT-free corn was cleaned-up by flash column chromatography (FCC) (3) using diethyl ether. The fraction which normally contains AFT is eluted off with 20% acetone in chloroform and brought to dryness

under N_2 . To this residue 200 μ l of benzene-acetonitrile (98+2) is added and then 5 μ l of this solution is spotted on a HPTLC plate and then over-spotted with various concentrations of AFT B₁ and B₂. The plate was then developed with either solvent A or solvent B.

RESULTS AND DISCUSSION

Corn oil contains 1.5 to 4.0% FFA (4). Assuming that none of the FFA are separated from the AFT during the clean-up procedure, then the final dilution used for spotting (here in our case 200 μ l for 1.0 g samples) would contain 0.75-2.0 μ g/ μ l of FFA. Thus a 5 μ l spot could contain 3.75-10.0 μ g of FFA; a concentration well within the range of interference.

Experiments with the isolated free fatty acid mixture showed that the R_f values are strongly concentration dependent (Table 1). The higher R_f 's are produced by the greater concentration of the FFA's presumably because the material occurring in the front of the spot deactivates the silica gel as migration proceeds and the following material encounters a less polar absorbant (2).

Attempts to exclude completely the FFA during the clean up procedure with FCC by mobile phase modification were unsuccessful. However it was found that a decrease in the amount of FFA was observed in the AFT fraction if acetone-chloroform mixtures (i.e. 20% A/C) were used to remove the AFT from the column. The CB clean-up procedure (5) which uses chloroform-methanol (97+3) to remove AFT from the clean-up column had significantly higher FFA in the AFT fraction than the FCC clean-up procedure as revealed by I₂.

TABLE 1
R_f Values of Corn Free Fatty Acids
of Differing Concentrations

Concentration of FFA ($\mu\text{g}/\text{spot}$)	R _f value ^a	R _f value ^b
1.4	0.35	0.49
3.7	0.35	0.49
7.4	0.39	0.50
22.0	0.44	0.54
36.6	0.48	0.56
73.3	0.54	0.60
220.0	0.63	0.68
366.5	0.66	0.69
1.5 ng - AFT B ₁	0.35	0.33
0.3 ng - AFT B ₂	0.26	0.28

^a Mobile phase: 8% acetone in chloroform.

^b Mobile phase: 8% acetone + 1% glacial acetic acid in chloroform.

However, the FCC clean-up procedure still gave recoveries of AFT greater than 100%. Using ammonia, diethylamine or other basic additives to both the column mobile phase and the TLC eluent to retard migration of the FFA were rejected because of the possibility of AFT degradation (6).

The problem was solved by the addition of 1% glacial acetic acid (GAA) to the TLC solvent. Mobile phase B caused the FFA to migrate significantly ahead of the AFT B₁ and B₂ with no overlapping and streaking with R_f values averaging 0.24 and 0.29 greater than AFT B₁ and B₂ respectively (Table 1). FFA in concentration ranges usually seen in AFT analysis gave an R_f

difference of 0.16 and 0.21 greater for AFT B₁ and B₂. The GAA addition also significantly reduced the FFA R_f differences due to concentration. Presumably the GAA causes complete protonation of the carboxylic acid portion of the FFA and eliminates any partial ionization which would contribute to the streaking. It also would decrease the polarity of the FFA thus giving them greater mobility through the silica gel and consequently a higher R_f.

Fluorescence enhancement by the FFA was also concentration dependent (Table 2). Even though higher concentrations of the FFA have a greater R_f difference compared to AFT-B₁ and B₂ as did the lower concentrations of FFA, they significantly enhanced the fluorescence to a greater degree than the lower concentrations of FFA. Apparently the tailing of the FFA in the higher concentrations was sufficient to over lay AFT B₁ and B₂. Using various concentrations of FFA's and over spotting with 1.5 ng of AFT B₁ and 0.3 ng B₂ mixture, the optimum concentration range of 1.33 μg to 7.33 μg of FFA was found to give the closest R_f's to AFT B₁ and B₂ using mobile phase A as HPTLC developing solvent (Table 1). The fluorescent enhancement produced at these concentrations averaged about 13.7% for AFT B₁ and 16.1% for AFT B₂. However larger concentrations of the FFA's (36.5 to 366.5 μg) produced an average of 35.7% enhancement for AFT B₁ and 28.6% for B₂.

Recovery results using actual corn extracts with a FCC clean-up procedure using a HPTLC solvent with and without GAA are depicted in Table 3. The average recovery without the GAA addition for B₁ and B₂ are 126.5% and 120.3%. With GAA addition to the mobile phase the average recovery for AFT B₁ is 98.75% and for AFT

TABLE 2

% Enhancement of Fluorescence* of 1.5 ng
AFT B₁ and 0.3 ng AFT B₂ by Corn FFA

FFA Concentration $\mu\text{g}/\text{spot}$	AFT - B ₁	AFT - B ₂
1.4	10%	14.3%
3.7	15%	20.4
7.4	16.2%	13.6
22.0	30.2%	22.9
36.6	33.9%	25.9
73.3	29.9%	24.0
220.0	39.5%	26.6
366.5	39.6	37.9

*Measured in triplicate using HPTLC plates.

TABLE 3

Recoveries* of AFT B₁ and B₂ in Spiked 1.0 g Corn Extracts

Spiked AFT B ₁ ,					
B ₂ in ppb	Recovered ^a ppb B ₁ , B ₂	Recovered ^b ppb B ₁ , B ₂			
12, 2.4	15.6, 2.8 (130, 117)	11.7, 2.3 (97.5, 95.8)			
20, 4.0	24.5, 4.3 (122.5, 107.5)	19.5, 3.9 (97.5, 97.5)			
60, 12.0	74.9, 13.4 (125, 111.7)	62.4, 12.2 (104, 101.7)			
100, 20.0	129.0, 29.4 (129, 147)	88.4, 13.7 (88.4, 68.5)			
200, 40.0	259.2, 46.2 (129.6, 115.5)	213, 34.7 (106.5, 86.7)			
500, 100	612.2, 123.0 (122.4, 123)	493, 99.3 (98.6, 99.3)			

*Measured in triplicate and averaged. % Recoveries in parenthesis.

^aMobile phase: 8% acetone in chloroform.

^bMobile phase: Chloroform, acetone, glacial acetic acid (92+8+1).

B₂ 91.6%. If the unexplained low value for the 20 ppb B₂ is dropped then the recovery averages 96.2.

For the AFT range used (12-500 ppb for B₁ and 2.4-100 ppb for B₂) an average difference of 27.75% for B₁ and 28.7% for B₂ in recovery was observed for the 2 different mobile phases. Obviously mobile phase B gave recoveries much closer to the actual values.

The prevention of the FFA interference in HPTLC analysis becomes even more significant for mold infected corn and older stored corn (1 yr old). This is because an increased level of FFA is observed due to the hydrolysis of corn triglycerides by fungi, bacteria, heat, seed damage, and moisture (3,7). In particular, high levels of FFA have been attributed to Aspergillus flavus contamination of cotton seed, coconut oil, and chocolate seed (8,9,10). These results should be interpreted only for the HPTLC plates and the developing solvent used. However an inspection of any developed TLC chromatograms used for corn AFT analysis with over-lapping I₂ positive spots could reveal FFA not removed during the clean-up procedure.

ACKNOWLEDGMENT

This work was supported by NIOSH Grant #2R01-OH-00796-02.

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